

Regulation of ligand-independent notch signal through intracellular trafficking

Kazuya Hori,¹ Anindya Sen,¹ Tom Kirchhausen^{1,2} and Spyros Artavanis-Tsakonas^{1,3,*}

¹Department of Cell Biology; Harvard Medical School; Boston, MA USA; ²Immune Disease Institute; Harvard Medical School; Boston, MA USA;

³Collège de France; Paris, France

Notch signaling is an evolutionarily conserved mechanism that defines a key cell fate control mechanism in metazoans. Notch signaling relies on the surface interaction between the notch receptor and membrane bound ligands in an apposing cell. In our recent study,²² we uncover a non-canonical receptor activation path that relies on a ligand-independent, intracellular activation of the receptor as it travels through the endosomal compartments. We found that notch receptor, targeted for degradation lysosomal degradation through multivesicular bodies (MVBs) is “diverted” toward activation upon mono-ubiquitination through a synergy between the ubiquitin ligase Deltex, the non-visual β -arrestin Kurtz and the ESCRT-III component Shrub. This activation path is not universal but appears to depend on the cellular context.

Regulation of Notch Trafficking

The notch pathway acts throughout development to link fate choices of a cell to those of its immediate cellular neighbors, ultimately affecting proliferation, apoptosis and differentiation.¹⁻³ notch malfunction has been associated with aberrant development in all metazoans and with various human diseases.⁴ The dosage of the notch signal defines an extraordinarily sensitive parameter that regulates the developmental outcome of signaling. Hence, cellular mechanisms controlling the dosage of activated notch receptors are of importance for the biology and pathobiology of the pathway.

Numerous studies, mostly in flies, implicated intracellular trafficking of notch receptor in both ligand-dependent and ligand-independent notch signaling.⁵⁻¹³ Endocytosis of notch may result in either down- or upregulation of signaling depending on how the receptor is sorted within the cell.^{14,15} Although these studies indicated an important role for intracellular trafficking in notch signaling, the compartments and molecular mechanisms underlying notch activation—once the receptor enters the endocytic path—remain unclear. Available evidence suggests that there may be several mechanisms that can lead to intracellular notch activation, either in a ligand-dependent or a ligand-independent manner. The process capable of modulating such intracellular notch signaling remains unknown, but ligand-independent activation of the receptor has been recently shown to be essential for the normal development of *Drosophila* blood cells.¹⁶

An important element of the endosomal sorting machinery is defined by the ESCRT (endosomal sorting complex required for transport) system.¹⁷ ESCRT is crucial in mediating the various steps leading to the sorting of membrane proteins into multivesicular bodies (MVBs) on their way to lysosomal degradation. It is also required for the morphogenesis of intraluminal vesicles and for the sorting of ubiquitinated cargo into these vesicles.¹⁷ As elaborated below, we identified an ESCRT member as a notch signal modifier. The ESCRT system consists of ESCRT-0, -I, -II, -III, and Vps4.¹⁷ ESCRT-0, -I, -II contain ubiquitin-binding domains, and are primarily

Keywords: notch signal, membrane trafficking, deltex, non-visual β -arrestin, ESCRT complex

Abbreviations: ESCRT, endosomal sorting complex required for transport; MVBs, multivesicular bodies

Submitted: 03/09/12

Accepted: 03/12/12

<http://dx.doi.org/10.4161/cib.19995>

*Correspondence to: Spyros Artavanis-Tsakonas;
Email: artavanis@hms.harvard.edu

Addendum to: Hori K, Sen A, Kirchhausen T, Artavanis-Tsakonas S. Synergy between the ESCRT-III complex and Deltex defines a ligand-independent Notch signal. *J Cell Biol* 2011; 195:1005–15; PMID:22162134; <http://dx.doi.org/10.1083/jcb.201104146>

involved in cargo sorting and in the recruitment and activation of ESCRT-III.¹⁷ The ESCRT-III subunits Vps20, Vps32 (Shrub), Vps24, Vps4 are assembled in this order, and contribute to the budding and scission of intraluminal vesicles into the vesicles.^{17,18} Vps4 is the ATPase required for ESCRT-III disassembly.¹⁷ Loss of ESCRT function modulates notch trafficking and leads to ectopic activation of notch signaling.^{8-12,19,20} The mechanisms underlying these events are complex, as different members of the ESCRT system exhibit distinct, non-overlapping phenotypic characteristics. For instance, the Vps22, Vps25, Vps36 members of ESCRT-II display non-identical mutant phenotypes involving notch activity, proliferation, or apoptotic resistance.²⁰ Moreover, there may be tissue specific functions of each component of ESCRTs.²⁰

Shrub is a Key Modulator of the Notch Signal Mediated by Deltex/Kurtz

We previously reported interactions between the non-visual β -arrestin Kurtz and the ubiquitin ligase Deltex leading to the regulation of notch trafficking, its degradation and consequential loss of signaling.²¹ We identified Shrub, a member of the ESCRT-III complex, as a major modulator of Deltex/Kurtz on notch signaling.²² Our genetic and molecular studies indicate that absence of Shrub suppresses the classic loss-of-notch wing-nicking phenotype associated with notch degradation due to the co-expression of Deltex and Kurtz.²² Use of Cut, a direct downstream target of notch signals, as a readout of notch activation revealed that its activation by Deltex can be suppressed by coexpression of Shrub. Expression of Shrub also amplifies the inhibiting effect of Kurtz on notch.²² But when Shrub activity is inhibited in a Deltex over-expressing background, we found a striking concomitant notch activation.²² This activation does not depend on the presence of the ligands for Delta or Serrate.²² In addition, biochemical analyses demonstrated that this activation was coupled with an accumulation of mono-ubiquitinated notch.²² ESCRT-III, through

Shrub, seems to play a distinct role in these phenomena, as ESCRT-I and -II components are not required for this Deltex-dependent notch activation.²² Consistent with this, mutants of ESCRT-I or ESCRT-II did not suppress the wing phenotype caused by the degradation of notch associated with the overexpression of Deltex and Kurtz.²² Therefore, our data suggest a check-point

by ESCRT-III and not ESCRT-I or -II in the ligand-independent and Deltex/Kurtz-dependent regulation of notch signaling.

Context Specific Regulation of the Notch Signal

A characteristic of the unconventional non-canonical activation of notch signaling we uncovered, is that it is context

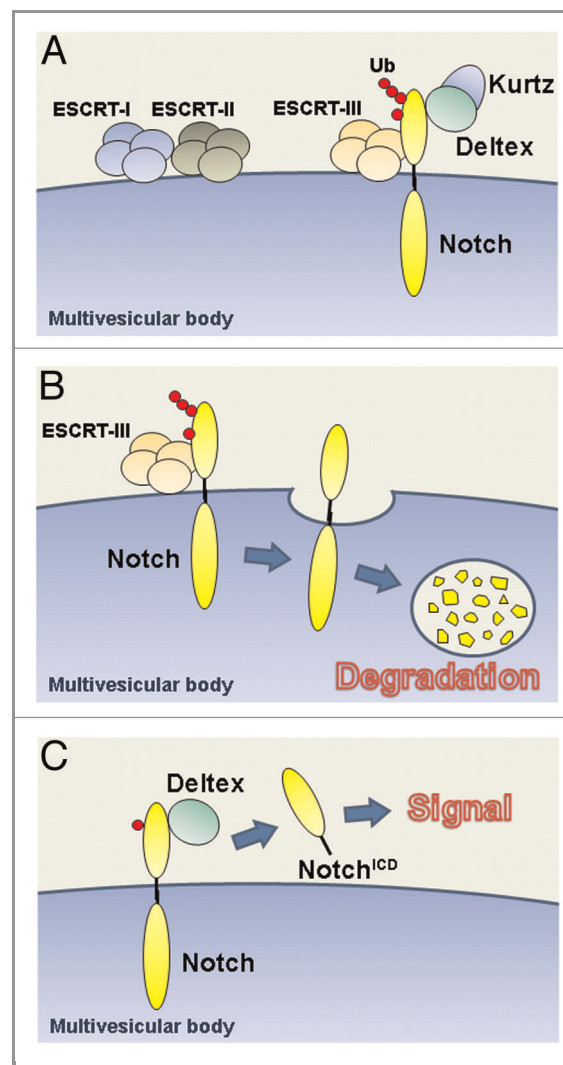


Figure 1. Shrub-Deltex-Kurtz dependent modulation of notch signaling. (A) The ubiquitination state of the notch receptor regulates its activation fate as it enters the endocytic path. While some steps in this path have been characterized, others simply illustrate our working hypothesis. (B) Our studies indicate that Deltex in synergy with Kurtz promotes the poly-ubiquitinated state of the receptor. This leads to the degradation of notch through the MVBs—a step regulated by Shrub that is a core component of the ESCRT-III complex. Our evidence is consistent with the notion that Shrub “surrounds” the ubiquitinated receptor—a role compatible with the previously suggested role of the yeast homolog Snf7. (C) The expression of Deltex, which physically interacts with notch, favors a mono-ubiquitinated state of the receptor and leads to a ligand-independent activation intracellular activation of notch (notch^{ICD}: the cleaved, activated form of notch).

dependent. In the wing disc for example, the notch signal induced by Deltex overexpression is seen mostly in the ventral region.^{6,22} Similarly, we found that the notch signal regulated by *deltex-shrub* is only manifested in the ventral region of the wing disc, indicating that this ligand-independent, intracellular activation of notch, depends on the cellular context.²² This context specificity is not associated with the glycosyltransferase Fringe (unpublished data), previously shown to regulate the differential activity of the notch receptor in the ventral vs. dorsal region of the wing disc.²³ However, treatment with the lysosomal inhibitor chloroquine abolishes this context specificity, suggesting that this effect depends on factors that regulate chloroquine sensitive lysosomal degradation.²² The mechanism underlying the involvement of lysosomal degradation in this context dependent

response is unclear but certainly noteworthy and the subject of future inquiries.

A Model for the Intracellular Activation of Notch

On the basis of these observations, we propose a model for *Shrub-Deltex-Kurtz* dependent modulation of notch activation (Fig. 1). *Shrub*, *Deltex*, and *Kurtz* regulate the trafficking of notch, and modulate the degradation of notch in the late-endosome/MVBs compartment. In the presence of *Shrub*, notch is sorted to the degradation pathway, resulting in downregulating the notch signal. In the absence of *Shrub* and/or *Kurtz*, *Deltex* promotes monoubiquitination, leading to notch activation.

Previous studies have indicated that there may be more than one mechanism that can potentially activate notch during its endosomal sorting.⁵⁻¹³ Our contribution

is based on linking notch activation to its ubiquitination state, modulated by the synergy of *Deltex*, *Kurtz* and *Shrub*. What remains unclear is the contribution of such a ligand-independent, context-dependent, intracellular activation in the normal biology and in the pathobiology of the notch signaling pathway.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Acknowledgments

This work was supported by NIH grants NS26084 and CA98402 (S.A.-T.), grant 07572 (T.K.), a JSPS Postdoctoral Fellowship for Research Abroad (K.H.) and a Postdoctoral Fellowship from the FSMA (A.S.). We would like to thank K. G. Guruharsha for critically reading the manuscript.

References

1. Artavanis-Tsakonas S, Rand MD, Lake RJ. Notch signaling: cell fate control and signal integration in development. *Science* 1999; 284:770-6; PMID:10221902; <http://dx.doi.org/10.1126/science.284.5415.770>
2. Schweisguth F. Regulation of notch signaling activity. *Curr Biol* 2004; 14:R129-38; PMID:14986688
3. Bray SJ. Notch signalling: a simple pathway becomes complex. *Nat Rev Mol Cell Biol* 2006; 7:678-89; PMID:16921404; <http://dx.doi.org/10.1038/nrm2009>
4. Louvi A, Artavanis-Tsakonas S. Notch and disease: A growing field. *Semin Cell Dev Biol* 2012; In press; PMID:22373641; <http://dx.doi.org/10.1016/j.semcdb.2012.02.005>
5. Coumalleu F, Fürthauer M, Knoblich JA, González-Gaitán M. Directional Delta and Notch trafficking in Sara endosomes during asymmetric cell division. *Nature* 2009; 458:1051-5; PMID:19295516; <http://dx.doi.org/10.1038/nature07854>
6. Hori K, Fostier M, Ito M, Fuwa TJ, Go MJ, Okano H, et al. *Drosophila* *deltex* mediates suppressor of Hairless-independent and late-endosomal activation of Notch signaling. *Development* 2004; 131:5527-37; PMID:15496440; <http://dx.doi.org/10.1242/dev.01448>
7. Wilkin MB, Carbery AM, Fostier M, Aslam H, Mazaleyra SL, Higgs J, et al. Regulation of notch endosomal sorting and signaling by *Drosophila* Nedd4 family proteins. *Curr Biol* 2004; 14:2237-44; PMID:15620650; <http://dx.doi.org/10.1016/j.cub.2004.11.030>
8. Vaccari T, Bilder D. The *Drosophila* tumor suppressor *vps25* prevents nonautonomous overproliferation by regulating notch trafficking. *Dev Cell* 2005; 9:687-98; PMID:16256743; <http://dx.doi.org/10.1016/j.devcel.2005.09.019>
9. Thompson BJ, Mathieu J, Sung HH, Loeser E, Rørth P, Cohen SM. Tumor suppressor properties of the ESCRT-II complex component *Vps25* in *Drosophila*. *Dev Cell* 2005; 9:711-20; PMID:16256745; <http://dx.doi.org/10.1016/j.devcel.2005.09.020>
10. Childress JL, Acar M, Tao C, Halder G. Lethal giant discs, a novel C2-domain protein, restricts notch activation during endocytosis. *Curr Biol* 2006; 16:2228-33; PMID:17088062; <http://dx.doi.org/10.1016/j.cub.2006.09.031>
11. Vaccari T, Lu H, Kanwar R, Fortini ME, Bilder D. Endosomal entry regulates Notch receptor activation in *Drosophila melanogaster*. *J Cell Biol* 2008; 180:755-62; PMID:18299346; <http://dx.doi.org/10.1083/jcb.200708127>
12. Vaccari T, Rusten TE, Menut L, Nezis IP, Brech A, Stenmark H, et al. Comparative analysis of ESCRT-I, ESCRT-II and ESCRT-III function in *Drosophila* by efficient isolation of ESCRT mutants. *J Cell Sci* 2009; 122:2413-23; PMID:19571114; <http://dx.doi.org/10.1242/jcs.046391>
13. Wilkin M, Tongngok P, Gensch N, Clemence S, Motoki M, Yamada K, et al. *Drosophila* HOPS and AP-3 complex genes are required for a *Deltex*-regulated activation of notch in the endosomal trafficking pathway. *Dev Cell* 2008; 15:762-72; PMID:19000840; <http://dx.doi.org/10.1016/j.devcel.2008.09.002>
14. Fortini ME. Notch signaling: the core pathway and its posttranslational regulation. *Dev Cell* 2009; 16:633-47; PMID:19460341; <http://dx.doi.org/10.1016/j.devcel.2009.03.010>
15. Yamamoto S, Charng WL, Bellen HJ. Endocytosis and intracellular trafficking of Notch and its ligands. *Curr Top Dev Biol* 2010; 92:165-200; PMID:20816395; [http://dx.doi.org/10.1016/S0070-2153\(10\)92005-X](http://dx.doi.org/10.1016/S0070-2153(10)92005-X)
16. Mukherjee T, Kim WS, Mandal L, Banerjee U. Interaction between Notch and Hif-alpha in development and survival of *Drosophila* blood cells. *Science* 2011; 332:1210-3; PMID:21636775; <http://dx.doi.org/10.1126/science.1199643>
17. Henne WM, Buchkovich NJ, Emr SD. The ESCRT pathway. *Dev Cell* 2011; 21:77-91; PMID:21763610; <http://dx.doi.org/10.1016/j.devcel.2011.05.015>
18. Wollert T, Wunder C, Lippincott-Schwartz J, Hurley JH. Membrane scission by the ESCRT-III complex. *Nature* 2009; 458:172-7; PMID:19234443; <http://dx.doi.org/10.1038/nature07836>
19. Moberg KH, Schelble S, Burdick SK, Hariharan IK. Mutations in *erupted*, the *Drosophila* ortholog of mammalian tumor susceptibility gene 101, elicit non-cell-autonomous overgrowth. *Dev Cell* 2005; 9:699-710; PMID:16256744; <http://dx.doi.org/10.1016/j.devcel.2005.09.018>
20. Herz HM, Woodfield SE, Chen Z, Bolduc C, Bergmann A. Common and distinct genetic properties of ESCRT-II components in *Drosophila*. *PLoS One* 2009; 4:e4165; PMID:19132102; <http://dx.doi.org/10.1371/journal.pone.0004165>
21. Mukherjee A, Veraksa A, Bauer A, Rosse C, Camonis J, Artavanis-Tsakonas S. Regulation of Notch signalling by non-visual β -arrestin. *Nat Cell Biol* 2005; 7:1191-201; PMID:16284625; <http://dx.doi.org/10.1038/ncb1327>
22. Hori K, Sen A, Kirchhausen T, Artavanis-Tsakonas S. Synergy between the ESCRT-III complex and *Deltex* defines a ligand-independent Notch signal. *J Cell Biol* 2011; 195:1005-15; PMID:22162134; <http://dx.doi.org/10.1083/jcb.201104146>
23. Fleming RJ, Gu Y, Hukriede NA. Serrate-mediated activation of Notch is specifically blocked by the product of the gene *fringe* in the dorsal compartment of the *Drosophila* wing imaginal disc. *Development* 1997; 124:2973-81; PMID:9247339